International Journal of Organic Agriculture Research and Development Vol. 2(2010).

Evaluation of sweet orange peel aqueous extract as root knot nematode suppressant

Abolusoro, S.A.^{1*}; Oyedunmade, E.E.A.² and Olabiyi, T.I.³

¹Department of Crop Production Technology, DAC/ABU Kabba, Kogi State, Nigeria ²Department of Crop Protection, University of Ilorin, P.M.B. 1515, Ilorin, Nigeria

ABSTRACT

The toxic effect of the aqueous extract of sweet orange (Citrus sinensis) to the root knot nematode, Meloidogyne incognita, was compared to that of carbofuran (a synthetic nematicide) in the laboratory and in the field during the 2007 and 2008 planting season at Ahmadu Bello University, College of Agriculture, Kabba. The laboratory experiment tested the effect of citrus peel extract and carbofuran on egg hatch and juvenile mortality of the nematode while the field experiment tested the effect of the treatments on soil and tomato root populations of the nematode. Sweet orange peel aqueous extract was applied at 0, 25, 50, 75 and 100% concentrations, while carbofuran was applied at 0, 250, 500, 750 and 1000ppm on the field and in the laboratory on egg hatch inhibition and juvenile mortality of M. incognita. The experiment lasted for a period of six months on the field and seven days in the laboratory for each year. The results from the field experiment showed that citrus peel aqueous extract and carbofuran solution brought about significant reduction in nematode multiplication rate and consequent root damage (root gall index) as compared with the untreated control. The higher concentrations (750 and 1000ppm) of sweet orange peel aqueous extract were significantly more effective than lower concentrations of 250 and 500ppm in suppressing nematode population in the soil and root. In the laboratory, citrus peel extract and carbofuran inhibited Meloidogyne incognita egg hatch and juvenile survival. The result indicates with respect to all the tested parameters that sweet orange peel aqueous extract is toxic to root-knot nematode and can be incorporated into its control system and as potential raw material for manufacturing organic based nematicide.

KEYWORDS: Peel of sweet orange, *Citrus sinensis*, root-knot nematode, carbofuran, tomato

Received: 12 December 2008 Accepted: 15 August 2010

 $*Correspondence \ author: \underline{steveabolusoro@yahoo.co.uk}$

³ Department of Crop and Environmental Protection, Faculty of Agricultural Sciences, Ladoke Akintola University of Technology, P.M.B. 4000, Ogbomoso, Nigeria

International Journal of Organic Agriculture Research and Development Vol. 2 (2010).

INTRODUCTION

MATERIALS AND METHODS

and Development Vol. 2 (2010).

International Journal of Organic Agriculture Research

Tomato crop is grown both as a perennial and an annual and it is herbaceous in nature. It is one of the most important vegetables in many regions of the world ranking second in importance to potato (Anonymous, 1989). It is a source of vitamins A, C and D. Its nutritional value cannot be over emphasized. Tomato is affected by wide range of pests. Prominent among them are the plant parasitic nematodes of which *Meloidogyne incognita* is identified as wide-spread and very devastating on tomato crop.

The disease cause by this group of nematode is the root knot disease and it's characterized by numerous pronounced swelling or gall on the root of tomato. Infected tomato plant suffers vascular damage and disturbed water and mineral uptake. Above ground symptoms are stunting, chlorosis and early senescence (Richard and Nicola, 1990). The most effective and most rapid control is achieved by the use of chemicals known as nematicides. However, nematicides are expensive and can be hazardous both within the crop and the soil. All these problems have greatly limited the use of chemical nematicide by many farmers in Nigeria (Egunjobi and Onayemi, 1981; Oyedunmade et al., 2009). The search for alternative control measures for root-knot nematode has thus become imperative. Hoan and Davide (1979), Maqbool et al (1987) and Alam et al (1980) suggested the use of plant extracts. Plant extracts are cheap and are readily available compared with conventional nematicides. They are environmentally safe (Oyedunmade et al., 1995; Zurren and Khan, 1984; Egunjobi and Onayemi, 1981). In an environmentally conscious world plant extract holds promise for their acceptability and use by resource constraint farmers.

Natural plant products are at present in the focus of research efforts because of their ability to produce environmentally less harmful, but efficacious bio-chemical substance (Schmutterer, 1990). This will greatly minimize the use of toxic synthetic chemicals. It is against this background that extract of the peel of sweet orange (*Citrus sinensis*) was tested for its toxicity on root knot nematode, *M. incognita*, since the toxicity of its component to other pests, especially the field insect pests, have been reported by researchers (Abolusoro, 2001; Olaifa *et al.*, 1987).

The study was therefore, conducted to (1) assess the effect of citrus peel extract on the egg hatch inhibition and on juvenile survival of M. incognita in the laboratory; and (2) investigate the effect of treating the root-knot nematode infested soil with citrus peel extract on the soil and root population of root-knot nematode M. incognita and the root damage of tomato growing in the medium.

2

The trial was conducted on a research field with known record of infestation of Meloidogyne incognita (Abolusoro, 2005) at Ahmadu Bello University, College of Agriculture Kabba, Kogi State, Nigeria between the month of July and December 2007 and was repeated at the same time in 2008. Tomato (cv. Roma V) seedlings were raised in sterilized nursery soil. The seedlings were transplanted to the permanent field on ridges 3 weeks after planting (3WAP) at a distance of 50 cm on ridges which were 90 cm apart. The experiment was 2 x 5 factorial fitted into randomized complete block design and each was replicated five times. Each plant was inoculated with 2000 eggs of M. incognita 6 WAP to increase the soil root knot nematode population of tomato plant to economic injury level. Fresh peel of sweet orange (C. sinensis) was collected; air dried and ground into fine powder; weighed on weighing balance and soaked in 1 litre of distilled water for 24 hours. The resultant filtrate was taken as the stock solution of strength 100%. Several dilution were made from the stock solution by adding the appropriate amount of distilled water to obtain 25, 50, 75% concentrations and 100ml of it was applied to each tomato plant at 7 WAP. Distilled water only served as the control (0%). Data were collected on number of nematodes in 200 g soil at planting and harvesting by method of Whitehead and Hemming (1965). The population of root knot nematode juveniles in 5g root was determined by the method described by Hooper (1990). The root damage, otherwise known as gall index, was determined using a rating scale 0-5 (Taylor and Sasser, 1988).

Citrus peel was subjected to phytochemical screening so as to determine the presence of the bioactive chemical components. The presence of flavonoids, tannin, saponins and alkaloids were determined by the method described by Trease and Evans (1989).

All data, except those on bioactive chemical components, were subjected to analysis of variance and means separated by Duncan multiple range test (Duncan, 1955).

LABORATORY EXPERIMENT

Egg hatch test: Eggs of root knot nematode, *Meloidogyne incognita*, were extracted using sodium hypochlorite (Hussey and Barker, 1973) from galled root raised in a pure culture of root knot nematode, *M.incognita*, race 2. One hundred freshly extracted root knot nematode eggs were introduced into each of the 40 Petri-dishes which were arranged in laboratory at room

International Journal of Organic Agriculture Research and Development Vol. 2 (2010).

temperature of $28 \pm 2^{\circ}$ C. Twenty milliliter (20ml) of each of the concentrations (100, 75, 50 and 25%) of the aqueous extract of the citrus peel was added separately to 100 freshly extracted root knot nematode eggs. Distilled water only (0% extract) served as the control. Every treatment was replicated eight times. Observations were made on egg hatch every 24 hour for 7 consecutive days. This was done by counting the number of second stage juvenile which emerged from egg using a stereomicroscope.

Juvenile mortality test: Ten milliliter of carbofuran solutions at 250, 500, 750 and 1000ppm concentrations and 25, 50, 75 and 100% aqueous extract of citrus peel were introduced into each of another set of 40 Petri-dishes. Distilled water only served as control (0%). Standardized 1ml juvenile suspension containing 100 juveniles of root knot nematode *M. incognita* was introduced into each of the transparent Petri-dishes containing different concentrations of citrus peel extract and carbofuran solution. The contents of the Petri-dishes were incubated at a temperature of $28\pm2^{\circ}$ C. The Petri-dishes were covered with glass to prevent evaporation. The experimental design in the laboratory was a randomized complete design. Each treatment was replicated 5 times. Counts of dead juveniles were made initially at 6 hours and then 12 hours of setting up the experiment. Thereafter, the observations were made at every 24th hour (daily basis) interval for a period of 7 consecutive days. The juveniles that did not respond to touch of needle were recorded as dead.

RESULTS

Table 1 shows the effect of citrus peel extract and carbofuran on soil and root population of *M incognita* as well as gall index of infected tomato. The result shows that the parameters under study were significantly lower in the carbofuran treatments as compared with the citrus peel aqueous extract. All the treatments reduced nematode population both in the soil and in the tomato root at harvest; and also tomato root damage (gall index) was significantly reduced in all the treatments as compared with the control. The level of reduction of the nematode populations in soil and tomato root, and gall index by carbofuran and citrus peel extract was directly proportional to the quantity of treatment applied.

Table 2 shows the comparative effect of carbofuran and citrus peel extract on egg-hatch inhibition of *Meloidogyne incognita*. The result shows a total egg-hatch inhibition at the higher concentrations of carbofuran and citrus peel extract from Day 1 to Day 7. Egg hatch inhibition was significantly

International Journal of Organic Agriculture Research and Development Vol. 2 (2010).

higher in all the treatments than the control. While few juvenile emerged at the lower level (25%) of citrus peel extract and 250ppm of carbofuran.

Table 3 shows the effect of carbofuran and citrus peel extract on juvenile mortality of M incognita under laboratory condition. The result shows that both carbofuran and citrus peel extract were effective in causing mortality on M. incognita juvenile. Mortality was observed at all the concentration levels of carbofuran and citrus peel treatment from 6^{th} hour of treatment application. 100% mortality was observed at all dosage levels of carbofuran 24 hours (Day 1) after treatment application. By day 2 of the experiment, 100% mortality was recorded in all the dosages of citrus peel treatments except at 25% concentration where 96% mortality was observed.

Table 1: Effect of citrus peel extract and carbofuran on soil and root population of M. Incognita as well as gall index

Off the infected tomato

2007 trial 2008 trial 2.90ab 2.88a 2.83a 4.25c 3.38ab 2.88a 2.88a 0.17 **2.**3a Gall index 2.88a 2.97a 2.83a 2.50c 3.13b 2.93a 2.97a 0.17 2008 trial Number o juvenile in 5g root 26.67e 18.33d 13.00c 12.33c 10.67b 29.33e 11.00b 12,00c 16.00c 6.67a 99.9 2007 trial 24.67e 17.33d 13.67c 12.00c 10.67c 27.00e 13.00c 11.67c 8,00b 5.67a 6.77 Multiplication rate (%) 4 PF/P1x100 2007 trial 2008 trial 2 1149b 43.81a 37.29a 30.72a 116.78b 36.90a 36.96a 466.7a 28.10a 21.85a 7.35 2007 trial 129.50b 117.1b 46.11a 41**.**81a 38.89a 30.49a 31.80a 26.95a 22.18a 36.64a 8.17 Population on (20cm) soil 2008 trial 2464.80h 896.51g 784.20e 648.21c p0'099 602.0b 459.0a 45.38 Final M. Incognita (PF) 2007 trial 877.21g 812.50e 644.90c 2458 00i 969 25h 2449.90i 774.00f 66p9,00d 565.71b 466.50a 21.94 2008 trial Initial M. Incognita 2101 2103 2110 2110 2114 2104 2097 2100 NS Population (P) 2007 trial 250ppm 2112 500ppm 2709 2098 2097 2099 2112 2099 2099 2103 NS 1000ppm 50% 75% 100% C. Sinensis 0 25% Concentration Cabofuran

Figures with the same letter (s) in the column do not differ significantly at P<0.05 according to

SE

Table 2: Comparative effect of carbofuran and citrus peel extracts on egg hatch inhibition of Meloidogyne incognita

	FIRST TRIAI	TRIAL				U	SECOND TRIAL) TRIAL							
Treatment	Concentration Day 1 Day 2 Day 3 Day 4 Day 5 Day 6 7Day	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	7Day	1Day	Day	Day	Day	Day	Day	Day
										2	3	4	5	9	7
C. Sinensis	25%	1.5a	1.5a	7.0b	9.0b	9.00	9.00	90 . 0	0a	1.4a	5.0a	7.2b	9.00	8.50	9.4
	20%	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
	75%	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
	100%	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
Carbofuran	250ppm	0a	1.5a	1.5a	1.5a	2.0b	2.0b	2.0b	0a	1.2a	1.3a	2.2a	2.2b	2.2b	2.2b
	%009	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
	750ppm	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
	1000ppm	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a	0a
Control	0	17.7b	33.0b	38.0c	62.0c 75.5d		91.0d	91.0d	16.9b	35b	41c	909	79d	93d	939
SE		1.03	2.12	1.65	1.23	1.44	1.55	0.55	1.04	2.12	1.70	1.23	1.42	0.51	0.55

Figure with the same letter in the column do not differ significantly at P < 0.05 according to Duncan's Multiple range test.

Ducan's multiple range test.

Table 3: Comparative effect of carbofuran and citrus peel extract on juvenile mortality of M. Incognita (J2) under . Laboratory condition

Table 4 shows the quantitative bioactive chemical compositions of sweet orange (citrus) peel. The table shows that citrus peel contains flavonoids, saponins and tannin while alkaloid was completely absent.

Table 4: Bio	-active chemical c	omposition of ci	trus peel
Alkaloid	Flavonoids	Saponins	Tannin
Nil	44.00%	9.78%	6.22%

DISCUSSION

The citrus peel extract and carbofuran significantly reduced the soil and tomato root population of the root knot nematode, M. incognita. Root damage (gall index) was also significantly reduced, in both treatments, in proportion to the levels of application and was significantly different from the control treatment. This observation agrees with those of earlier researchers. Abid et al (1995) and Sellami and Moufarrah (1994) reported the effectiveness of various plant extract used in their various experiments in suppressing both plant root and soil population build up of nematode and consequent reduction in the root damage in the tested crops. The citrus peel extract brought about egg hatch inhibition as well as heavy mortality of nematode juvenile and compared favourably with synthetic nematicide (carbofuran). The observed nematoxic effects of the citrus peel extract can be attributed to the presence of nematicidal chemical components that were seriously injurious to Meloidogyne incognita eggs and infective second stage juveniles (J2). The findings from the study corroborate that of earlier researchers like Pandy (1990) and Mani and Al Hinai (1998) who reported the toxicity of various plant extracts used in the experiments.

Saponins, flavonoids and tannin were present in the citrus peel and are probably responsible for the toxicity of the citrus peel extracts to M. incognita. This observation agrees with Alam et al (1989) who reported the effectiveness of flavonoid, tannin and saponin in reducing M. incognita population and enhancing egg-hatch inhibition of M. incognita in their various experiments. This type of control on plant parasitic nematode had been reported by many workers including Oyedunmade et al (2001), Oyedunmade (2004) and Olabiyi and Oyedunmade (2004). The exhibited nematicidal properties may be due to the presence of bioactive chemical component including Saponin, flavonoid and tannin in the citrus peel extracts (Olabiyi et al., 2008).

The significance of this study thus underpin the potency of citrus peel extracts as a viable alternative to synthetic nematicide in the control of nematode pests on susceptible crops. The Agro-chemical companies can

וובמווובווו			2		ہے							วี	SECOND INAL		Ļ				
Cc Tra	Concen- Tration (%)	6hrs	12hrs	Day1	Day2	Day3	Day4	Day5	Day6	Day 6 7	6hrs 1	12hrs	Day1	Day2	Day3	Day4	Day5	Day6	Day 7
Citrus Sinensis	25%	J0	10.5f	71.0c		96.0b 100a	100a	100a	100a	100a 0e		10.5d	10.5c	100a	100a	100a	100a	100a	100a
	%09	1.0e	19.0e	87.0b	100a	100a	100a	100a	100a	100a	5.7d	19.1cd	72.2b	100a	100a	100a	100a	100a	100a
	75%	10.0d	24.0d	100a	100a	100a	100a	100a	100a	100a 16.0c		25.0c	81.5b	100a	100a	100a	100a	100a	100a
	100%	20.0b	40.5c	100a	100a	100a	100a	100a	100a	100a	20.0b 41.2b		100a	100a	100a	100a	100a	100a	100a
Carbofuran	250ppm	8.0d	27.0d 100a	100a	100a	100a	100a	100a	100a	100a	8.5d	26.1c	100a	100a	100a	100a	100a	100a	100a
	500ppm	16.2c	36.0c	100a	100a	100a	100a	100a	100a	100a 16.3c		37.0b	100a	100a	100a	100a	100a	100a	100a
	750ppm	25.0b	52.5b	100a	100a	100a	100a	100a	100a	100a 16.3c		37.0b	100a	100a	100a	100a	100a	100a	100a
	1000ppm	32.0a	60.0a	100a	100a	100a	100a	100a	100a	100a 3	34.0a	62.0a	100a	100a	100a	100a	100a	100a	100a
Control	0	JO	0g	po	00	qo	3.3b	90.9	8.5b	10.1b 0e		0e	P0	qo	q0	qo	qo	q ₀	qo

Figures followed by the same letter(s) in the same column are not significantly different at P<0.05 using

International Journal of Organic Agriculture Research and Development Vol. 2 (2010).

start extracting the bioactive components of the citrus peel as raw materials for manufacturing of botanical based nematicide as a guarantee for environmental safety and production of safe to eat food.

REFERENCES

- Abid, M; M.A. Zaki and M.A. Maqbool (1995). Neem derivates for the control of root knot nematode, M. javanica, on okra. Pakistan J. of Phytopath., 7(2): 212-214.
- Abolusoro, S.A. (2001). Efficacy of some plant materials for the control of cowpea bruchid *Callosobruchus maculata* (Fab). Journal of Agric. Technology, **9**: 1-6.
- Abolusoro, S.A. (2005). Nematicidal activities of some selected botanicals on a root-knot nematode affecting tomato, *Lycopersicon esculentum* (L.) Mill, Ph.D Thesis submitted to the Department of Crop Production, University of Ilorin, Ilorin, Nigeria: 206pp.
- Alam, M.M.; M. Ahmed and A.M. Khan (1989). Effect of organic amendment on the growth and chemical composition of tomato, egg plant and chilli and their susceptibility to attack by *Meloidogyne incognita*. Plant and Soil, **57**: 231-236.
- Anonymous (1989). F.A.O. Production Year Book, volume 42, Rome, Italy.
- Duncan, D.B. (1955). Multiple range and multiple F tests Biometrics 11, 1-42.
- Egunjobi, A.O. and S.O. Onayemi (1981). The efficacy of water extracts of neem (*Azadirachta indica* A. Juss.) leaves as systematic nematicides. Nigeria Journal of Plant Protection, **5**: 70-74.
- Harborne, J.B. (1973). Phytochemical methods: A guide to modern techniques of plant analysis. Halsted Press, John Wiley and sons inc, New York, 278pp.
- Hoan, L.T. and R.G. David (1979). Nematicidal properties of root extracts of seventeen plant species on *Meloidogyne incognita*. Phil. Agric **62**: 285-295
- Hooper, D.J. (1990). Extraction and processing of plant and soil nematodes. In: Plant parasitic nematodes in subtropical and tropical agriculture. Eds. M. Luc, R.A. Sikora and J. Bridge. CAB International: 45-68.
- Hussey, R.S. and K.R. Barker (1973). A comparison of method collecting inocula for *Meloidogyne* species including a new technique. Plant Disease Report, **57**: 1025-1028.

10

International Journal of Organic Agriculture Research and Development Vol. 2 (2010).

- Mani, A. and M.S. Al Hinai (1998). Toxicity of Harnal, *Rhazya stricta*, to *Meloidogyne incognita* and *Paratylenchus jordanenes*. Nematol. Medit. **26**: 27-30.
- Maqbool, M.; A.S. Hashmi and A. Oghaffar (1987). Effect of latex extracts from *Euphorbia caducifola* and *Calotropis* procera on root-knot nematode *M. incognita* infested tomato and egg plant. Park J. Nematol, **5**: 43-48.
- Olaifa J.I.; W.O. Erhun and A.E. Akingbohungbo (1987). Insecticidal activities of some Nigeria plants, Insect Sciences and Appl. **8(2)**: 221-224
- Olabiyi, T.I.; E.E.A. Oyedunmade and G.T. Ibikunle (2008). Phytochemical screening and nematoxic effect of brimstone, *Moringa lucida*, on nematode pests of Amaranth, *Celosia argentea*. Biological Agriculture and Horticulture, **26**: 131-137.
- Oyedunmade, E.E.A (2004). Laboratory and field toxicities of the African marigold (*Tagetes erecta*) to root-knot nematodes. The Plant Scientist, 4: 115-121.
- Oyedunmade, E.E.A. and T.I. Olabiyi (2004). In-vitro and in-vivo toxic effects of bitter leaf, *Vernonia amygdalina* Del to the root nematode *Meloidogyne incognita*. Nigeria Society of Experimental Biology Journal, 4: 15-19.
- Oyedunmade, E.E.A.; S.A. Abolusoro and T.I. Olabiyi (2009). Nematicidal activities of carbofuran and some organic materials on plant parasitic nematode control on tomato. International Journal of Nematology, **19(1)**. 96-102
- Oyedunmade, E.E.A; O.F. Ayo-Oke and T.I. Olabiyi (2001). Comparative efficacies of soil amendment and a synthetic nematicide on growth, yield and food components of a root-knot infected soyabean, *Glycine max*. (L). Merril. Nigerian Society of Experimental Biology Journal, 1: 367-371.
- Oyedunmade, E.E.A.; J.O. Babatola and T.I. Olabiyi (1995). The effects of two crop residues on nematode in cowpea cultivation. Nematol. Medit., 23: 61-64.
- Pandy, R.L. (1990). Studies on the phytonematoxic properties of extracts of some medicinal and Aromatic plants. International Nematology Network Newsletter, 7: 19-20.
- Richard, A.S. and G. Nicola (1990). Nematode parasite of food legumes. Plant parasitic nematode in sub-tropical and tropical agriculture. Eds. Luc. M; Sikora, R.A. and J. Bridge. CAB International, 197-198.
- Schmutterer, H. (1990). Properties and potentials of natural pesticide from the neem tree, *Azadirachta indica*. Ann. Rev. Entomol. **35**: 271-298.

- Sellami, S. and A. Mouffarah (1994). Effects of some aqueous plant extracts on juvenile hatching and larval mortality against *M. incognita* Mededelingen Faculteit-Land bouwkundige-en Toege paste Biologi sche-wetensch appen, Universiteit-gent., **59(26)**: 813-816.
- Taylor, A.L. and J.N. Sasser (1978). Biology, identification and control of root knot nematode, Meloidogyne species. North Carolina University Graphic Press. 111pp.
- Trease, E.B. and W.A. Evans (1989). Preliminary screening of plants for their chemical constituents, CAB International, 104pp.
- Whitehead, A.G. and J.R. Hemming (1965). A comparison of some quantitative methods of extracting small vermiform nematodes from soil. Ann. Appl. Bio. **55**: 25-38.
- Zurren, S. and M.I. Khan (1984). Nematicidal activity in some plant lattices. Pak J. Nematol., **2(2)**: 67-77. Proceedings of the VIIth International Rangelands Congress, 26 July August 2003, Durban, South Africa, pp. 543556.