

Chemical Composition of Biological Pesticides in Leaf of *Chromolaena odorata*

^{*1}Godonu Kolawole Gbemavo and ²Feyisola Roseline Tolulope

¹Department of Crop Science, College of Agriculture, Lagos State University of Science and Technology Ikorodu, Lagos State.

²Department of Plant Science, Olabisi Onabanjo University, P.M.B. 2002, Ago Iwoye Ogun state

ABSTRACT

The increased use of pesticides in the control of vector pests has raised some concern over the adverse effect of these chemicals on humans, domestic animals and their environment. This research was undertaken to harness the beneficial use of the extract of *Chromolaena odorata*, as biopesticide.

This study aims to determine bio-pesticidal chemical composition in ethanolic extract of *Chromolaena odorata* (Siam weed). Fresh and healthy leaves of *Chromolaena odorata*, were harvested. They were air dried for four weeks on the laboratory bench, pulverized and extracted using soxhlet apparatus and ethanol as the solvent. The bioactive of ethanolic extracts from the siam weed were profiled using gas chromatography flame ionization detector (GC-FID). The ethanolic extracts underwent GC-FID analysis, which identified 49 components. According to the bioactive contents of the extracts from the plant, these substances fall within the class of terpenes, esters, sterols, and triterpenoids, which were recognized to have active plant-protective and health-improving effects on economic food crops. Consequently, from the various literatures, the GC-FID analysis of the phytochemical components found in the siam weed extracts showed the potential to serve as biological insecticides, nematocides, bactericides and fungicides.

Keywords: *Chromolaena odorata*; bio-pesticides; bio-active; phytochemical

INTRODUCTION

Chromolaena odorata, (Siam weed), is a scrambling perennial plant that reaches a height of 2-3 cm. Its stems are straight, pithy, brittle, and easily branch. The leaves have an arrowhead shape and measure 6–12 cm in length and 3–7 cm in breadth. They have three veins that resemble a pitchfork (Pandurangan *et al.*, 2015).

Native to Central and South America, *C. odorata* is a weedy herb that has spread throughout tropical and subtropical regions (Gautier, 1992; Omokhua *et al.*, 2016). As a plantation cover crop, it was initially brought to Southeast Asia in the 1920s and Africa in 1940. Since then, it has proliferated all over the world (Rouw, 1991; Kouame *et al.*, 2013). For its several therapeutic benefits, *C. odorata* has been used historically. It is particularly useful externally for conditions including inflammation, skin infections, and wound healing. Research has shown that the leaf extract possesses anti-inflammatory, (Owoyele *et al.*, 2005; Pandith *et al.*, 2013), anti-bacterial, (Suksamrarn *et al.*, 2004; Stanley *et al.*, 2014) anti-cancer (Kouame *et al.*, 2013; Adedapo *et al.*, 2016), anti-convulsant (Amazuet *et al.*, 2013) anti-diabetic (Onkaramurthy *et al.*, 2013; Uhegbu *et al.*, 2016), anti-fungal (Ngono *et al.*, 2006; Naidoo *et al.*, 2011) antioxidant (Vijayaraghavan *et al.*, 2013; Boudjeko *et al.*, 2015), and anti-parasitic (Thoden *et al.*, 2007; Ezenyi *et al.*, 2014), hemostatic and wound healing (Pandurangan *et al.*, 2015;

Owoyele *et al.*, 2005; Pandith *et al.*, 2013; Pandith *et al.*, 2013), and hepato-protective activities (Alisi *et al.* 2011; Asomugha *et al.*, 2014).

The antioxidant properties of the plant help to preserve the development of keratinocytes and fibroblasts on the wounds, which is why mending wounds is so effective (Owoyele *et al.*, 2005). In Vietnam, *C. odorata* is widely used for traditional wound healing. The aqueous extract from the leaves has also been used to treat skin infections and soft tissue burns (Thang *et al.*, 2001; Phan *et al.*, 2001). Report has it that *Chromolaena odorata* has been used as bio-pesticide against beans weevils

Essential oils from the plant have shown anti-fungal and anti-microbial activities and anti-radical potential (Félicien *et al.*, 2012), mortality of the maize grain weevil, *Sitophilus zeamais* (Bouda *et al.*, 2001), in vitro potential cytotoxic activity on human epidemic cell line (Koba *et al.*, 2011), toxic on *Rhipicephalus lunulatus*, ectoparasite of the dwarf goat (Pamo *et al.*, 2004), exploited as insecticide (Noudogbessiet *et al.*, 2008), ovicide and larvicide (Félicien *et al.*, 2012). The chemical constituents of its volatile oils are always dominated by ubiquitous mono- and sesquiterpenes of diverse structural pattern. Some major compounds of its oils so defined include β -caryophyllene, germacrene-D, bicyclgermacrene, geijerene, (Z)- β -farnesene, α -pinene, pregeijerene, camphor and limonene (Owolabiet *et al.*, 2010;

Kossouhet *al.*, 2011; Suwaibahet *al.*, 2012; Joshi 2013). This research falls within the economic pillars under the good health and well-being of the vision 2030 Sustainable Development Goals. The research provides foundational background information on the production of biological pesticides, safe to eat foods for wellness and healthy life. Thus, the aim of this study is to assess the chemicals composition of biological pesticide in Siam weed leaf (*Chromolaena odorata*).

MATERIALS AND METHODS

Plant Material

Fresh *Chromolaena odorata* were harvested within the premises of Lagos State University of Technology, Ikorodu and were identified by a botanist. The leaves were carefully removed from the stem and washed in running water to remove contaminants. They were air dried at room temperature in an open laboratory space for four weeks and pulverized into powder.

Preparation of Extracts

The extraction was done using soxhlet apparatus and ethanol as the solvent according to the method described by Airaodion *et al.*, (2019). 25 g of pulverized leaves of the three species were packed into the thimble of the soxhlet extractor and 250 mL of ethanol was added to a round bottom flask, which was attached to the Soxhlet extractor and condenser on a heating mantle. The solvent was heated using the heating mantle and began to evaporate moving through the apparatus to the condenser. The condensate dripped into the reservoir housing the thimble containing the sample. Once the level of the solvent reached the siphon, it poured back into the round bottom flask and the cycle began again. The process was allowed to run for a total of 18 hours. Once the process was completed, the ethanol was evaporated in a rotary evaporator at 35°C with a yield of 2.55 g which represents a percentage yield of 10.20%. The extract was preserved in the refrigerator until when needed.

Gas Chromatography (GC) Analyses

Siam weed extract (200 ml) was taken to Central Research laboratory, Federal University of Technology, Akure, Ondo State, Nigeria for

bioactive constituents' analysis. The chemical composition of the crude extract was performed using gas chromatography coupled with flame ionization detector (GC/FID). The GC analysis was performed using a chromatography equipped with a flame ionization detector (FID) and two capillary columns of different polarities OV type: 101 (25 m x 0.22 mm x 0.25 µm) and Carbowax 20 M (25 m x 0.22 mm x 0.25 µm). The carrier gas was Helium with a flow rate of 0.8 ml/min and the oven programming temperature was between 50 and 200°C with a gradient of 5°C/min. CPG/MS coupling was performed on a DB1-type fused silica capillary column (25 m x 0.23 mm x 0.25 µm) with Helium as a carrier gas and temperature programming identical to that of the GC (AOAC, 2019).

Identification of the Components

Bioactive compounds extracted from the plant extracts were identified on the basis of GC retention time (RI, determined with reference to homologous series of n-alkanes C8–C25, under identical experimental conditions) on HP-5MS column and matching of the spectra with computer software data of standards (Replib and Mainlab data of GC–MS systems).

RESULTS

GC-FID chromatogram of extracts revealed that forty-nine bioactive compounds are present in *Chromolaenaodorata* (Siam weed). The GC showed the concentrations of the various compounds as a function of retention time while the peak height showed the relative concentrations of the components present in the extract, while the compounds were identified through flame ionization detector attached with the gas chromatography. The identified compounds with their name, % peak area, and retention time were categorized and summarized into five chemical groupsterpenes, triterpenoids, fatty acid, hydrocarbons, esters.

The yield of the extracts obtained from the soxhletethanolic extracts of the leaves of *C. odorata* was 10.20% calculated on a dry weight basis, Table 1 presents the components identified, their percentage compositions and retention indices based on their elution on the HP-5MS column. A total forty-nine compounds were characterized and identified by GC-FID.

Table 1: The Chemical Compositions of *Chromolaenaodorata*

S/N	RT	%	AREALIBRARY ID	REF#	CAS#	QUAL	3	7.117	1.40	4-Acetoxybenzaldehyde	35587	000878-00-2	42
										2,4,6-Cycloheptatrien-1-one,2-hydroxy-	10091	000533-75-5	30
1	4.946	0.47	Catechol	5860	000120-80-9	59				4-Acetoxybenzaldehyde			
			1H-Benzotriazole	9260	000095-14-7	53					35588	000878-00-2	27
			Catechol	5859	000120-80-9	50	4	7.413	0.77	Copaene	68472	003856-25-5	96
										alfa-Copaene	68522		96
2	5.354	3.68	5-Hydroxymethylfurfural	11339	000067-47-0	72				Copaene	68473	003856-25-5	96
			5-Hydroxymethylfurfural	11338	000067-47-0	64							
			1H-Pyrazole, 4,5-dihydro-3- methyl-1-propyl	11489	026964-49-8	47							

5	7.457	0.91	Copaene	68472	003856-25-5	96	24	15.234	0.99	Pentadecanoic acid	104278	001002-84-2	83
			Copaene	68473	003856-25-5	96				Cyclopentadecane	74572	000295-48-7	45
			Copaene	68474	003856-25-5	95				Pentadecanoic acid	104282	001002-84-2	42
6	8.087	4.18	Benzoic acid, 4-methoxy-	26627	000100-09-4	93	25	15.301	0.39	3,4-Dimethyl-2-prop-2-enyl-2,5 dihydrothiophene 1,1-dioxide	53481	1000305-61-1	70
			Benzoic acid, 4-methoxy-	26636	000100-09-4	93						1000159-38-5	
			Benzoic acid, 4-methoxy-	26632	000100-09-4	90							
7	11.442	2.10	3-[(Z)-1-Butenyl]-4 vinylcyclopentene	23445	052886-04-1	35				Cycloheptane, 4-methylene-1-methyl methyl-1-propen-1-yl)-1-vinyl-	68765		66
			6-(1'-Oxo-2'-propenyl)-1,3-cis,cis cyclooctadiene	33411	138146-05-1	35						1000131-71-1	
			Spiro[4.4]nona-1,6-diene, (S)							trans-Z.-alpha.-Bisabolene epoxide	83578		62
				9618	039746-39-9	35							
8	11.672	2.49	Bicyclo[4.1.0]heptane,-3-cyclopropyl,-7-hydroxymethyl, trans	36538	1000223-00-6	45	26	15.375	0.48	p-Menth-8(10)-en-9-ol, cis- 1,3,3-Trimethylcyclohex-1-ene-4-carboxaldehyde, (+,-)-	27588	015714-13-3	55
			Bicyclo[6.1.0]nonane, 9-(1-methyle thylidene)							13-Tetradec-11-yn-1-o	72625	1000131-00-4	50
			2,3-Diazabicyclo[2.2.1]hept-2-ene,5-ethenyl-4,7,7-trimethyl-,(1.alpha.,4.alpha.,5.beta.)-	35103	056666-90-1	43	27	15.405	0.33	1H-3a,7-Methanoazulene, octahydro-1,4,9,9-tetramethyl-	70783	025491-20-7	50
				34882	1000221-84-7	38				3,4-Dimethyl-2-prop-2-enyl-2,5 dihydrothiophene 1,1-dioxide	53481	1000305-61-1	41
9	12.242	1.33	Phenol, 3,4,5-trimethoxy-	51859	000642-71-7	95				trans-Z.-alpha.-Bisabolene epoxide	83578	1000131-71-1	38
			3,4,5-Trimethoxyphenol	51857	030225-76-4	94							
			Phenol, 3,4,5-trimethoxy-	51860	000642-71-7	72	28	15.464	1.08	Caryophyllene oxide	83535	001139-30-6	78
10	12.686	2.48	1,2,3-Trimethoxybenzene	38847	000634-36-6	38				1H-3a,7-Methanoazulene, octahydro 1,4,9,9-tetramethyl-	70783	025491-20-7	55
			1,2,3-Trimethoxybenzene	38845	000634-36-6	30				Diazoprogersterone	195535	1000255-30-9	52
			Vanillic acid	38762	000121-34-6	30							
11	12.871	2.14	Octacosylpentyl ether	263355	1000406-42-4	27	29	15.523	0.56	Alloaromadendrene	68577	025246-27-9	81
			Hexacosylpentyl ether	257324	1000406-42-3	27				(-)-Isolongifolol, methyl ether	98688		
			Carbonic acid, eicosyl vinyl ester	219345	1000382-54-3	25				9-Isopropyl-1-methyl-2-methylene-5oxatricyclo[5.4.0.0(3,8)]undecane		1000333-80-8	64
12	13.257	1.72	Cyclohexene, 1-nitro-	12014	002562-37-0	38					83636	1000185-86-6	52
			2-Heptyne	2867	001119-65-9	38							
			2-Heptyne	2864	001119-65-9	38							
13	13.516	1.20	Benzenepropanoic acid, 4-methoxy, beta-oxo-, methyl ester	72048	1000338-13-0	93	30	15.627	3.47	Bicyclo[2.2.1]heptane-2,5-dione,1 trimethyl-	36330	004230-32-4	50
			4'-Methoxybutyrophenone	45847	004160-51-4	87				2,4-Dimethyl-3-nitrobicyclo[3.2.1] octan-8-one	62339	1000192-17-3	47
			4'-Methoxybutyrophenone	45850	004160-51-4	87				2([H]-Naphthalenone, octahydro-4a, 7,7-trimethyl-, trans-	59812	054699-31-9	47
14	14.064	1.12	6-Hydroxy-4,7-dimethoxy-2H-1,3-be zodioxole-5-carbaldehyde	88870	849798-24-9	53	31	15.679	0.92	2([H]-Naphthalenone, octahydro 4a, 7,7-trimethyl-, trans-	59812	054699-31-9	81
			Tetramethoxybenzaldehyde	89005	065884-12-0	46				Fumaric acid, 2-chloropropyl penta decyl ester	239635	1000348-57-2	64
			Piperonylic acid, 5-nitro-	75528	000716-32-5	30				Hexacosyl acetate	248621	000120-80-9	60
15	14.101	1.21	6-Tridecene, 7-methyl-	61879	024949-42-6	70							
			7-Tetradecene, (Z)-	61868	041446-60-0	51							
			6-Tetradecene, (E)-	61859	041446-64-4	51							
16	14.219	0.97	4-((1E)-3-Hydroxy-1-propenyl)-2-methoxyphenol	47528	1000297-95-5	95	32	15.760	0.92	Octacosylheptafluorobutyrate	273239	1000351-83-6	87
			2-Methylthianaphthene-1,1 dioxide	48558	006224-55-1	64				cis-11-Hexadecenal	100562	053939-28-9	64
			(E)-4-(3-Hydroxyprop-1-en-1-yl)-2 methoxyphenol	47541	032811-40-8	58						195194-80-0	
17	14.419	2.31	Tetradecanoic acid	91415	000544-63-8	98	33	15.979	1.93	2-Piperidinone, N-[4-bromo-n-butyl]-	95915		64
			Tetradecanoic acid	91420	000544-63-8	97				5,6,6-Trimethyl-5-(3-oxobut-1-enyl)-1-oxaspiro[2.5]octan-4-one	98357	1000192-73-9	70
			Tetradecanoic acid	91418	000544-63-8	97				17-Pentatriacotene	265112	006971-40-0	
18	14.553	1.54	Pyridine, 4-methyl-, 1-oxide	5680	000544-63-8	43				2-(2R,4aR,8aS)-4a-Methyl-8-methyl enedecahydronaphthalen-2-yl)acrylaldehyde	81710	003650-40-6	55
			Silane, chlorotriethyl-										
			1H-3a,7-Methanoazulene, octahydro- 1,4,9,9-tetramethyl-	24908	000544-63-8	38				n-Hexadecanoic acid	117418	000057-10-3	99
				70783	000544-63-8	35	34			n-Hexadecanoic acid	117419	000057-10-3	99
19	14.686	0.82	2-Cyclopenten-1-one, 4-hydroxy-3-methyl-(2-propenyl)-	26892	000551-45-1	53	35	15.960	2.07	Octadecanoic acid	144271	000057-11-4	90
			Cyclopentaneacetaldehyde, 2-formyl -3-methyl-alpha-methylene-	36381	005951-57-5	50				Nonadecane, 1-chloro-	161769	062016-76-6	89
			Carane, 4,5-epoxy-, trans	25896	006909-20-2	49	36	15.997	1.42	Hexadecane	89844	000544-76-3	89
20	14.753	0.48	5-Ethyl-2-furaldehyde.	10622	023074-10-4	58				Carbonic acid, eicosyl vinyl ester	219345	1000382-54-3	55
			3-Cyclohexene-1-carboxaldehyde, 1-methyl-	10785	000931-96-4	52	37	16.049	2.93	Nonadecane, 1-chloro-	161769	062016-76-6	93
			1.alpha., 4a.beta., 8a.alpha.-Deca hydro-1-naphthalenol	27803	036159-47-4	49				1-Bromodocosane	232144	006938-66-5	50
21	14.901	1.57	4-(2,2-Dimethyl-6-methylenecyclohe xy)butanal	59779	095452-13-4					Carbonic acid, eicosyl vinyl ester	26632	1000382-54-3	46
			2([H]-Naphthalenone, octahydro-1,4 a-dimethyl-(1.alpha.,4a.beta.,8a.alpha.)-	47962	022738-31-4	45	38	16.545	0.99	Hexadecanoic acid, ethyl ester	144307	000628-97-7	97
			2-Decanoic acid	37828	001851-90-7	43				Hexadecanoic acid, ethyl ester	144309	000628-97-7	95
22	15.071	1.41	Tridecanedial	76252	063521-76-6	70	39	16.716	1.53	Undecanoic acid, ethyl ester	78053	000627-90-7	93
			Bicyclo[3.1.1]heptane, 2,6,6-trimethyl-, (1.alpha.,2.beta.,5.alpha.)	17424	006876-13-7	53	40	16.841	1.80	Cholest-3-en-2-one, 2-methylene-, 239014		022599-96-8	70
			Dodeca-1,6-dien-12-ol, 6,10-dimethyl-	74503	1000156-13-8	42				Cyclopentadecane	74572	000295-48-7	56
23	15.123	4.43	2-Pentadecanone, 6,10,14-trimethyl	128827	000502-69-2	91	41	16.893	1.60	tert-Hexadecanethiol	119379	025360-09-2	55
			2-Pentadecanone, 6,10,14-trimethyl	128823	000502-69-2	90				Methyl pentadecyl ether	104447	007307-52-0	53
			2-Pentadecanone, 6,10,14-trimethyl	128826	000502-69-2	90				Phytol	155849	000150-86-7	52
										Octacosylpentyl ether	263355	1000406-42-4	49
										9,12-Octadecadienoic acid (Z,Z)-	140138	000060-33-3	48
										Linoleic acid	140124	000506-21-8	97
										9,12-Octadecadienoic acid (Z,Z)-	140139	000060-33-3	96
										Cyclohexanol, 3-ethenyl-3-methyl-2-(1-methylethenyl)-6-(1-methylethyl),[1R(1.alpha.,2.alpha.,3.beta.,6.alp	85869	035727-45-8	90

		lin ...)]			
		Pentadecafluorooctanoic acid, octa			
		decyl ester	274630	1000406-04-8	49
		Nonadecyltrifluoroacetate	227165	1000351-76-3	49
42	16.997 2.33	Linoleic acid ethyl ester	167367	000544-35-4	89
		Z,Z-4,15-Octadecadien-1-ol acetate	167375	086252-67-7	83
		6-Tridecane	48000	042371-66-4	83
43	17.019 1.16	13-Octadecenal, (Z)-	126830	058594-45-9	60
		Oxirane, tridecyl-	89784	018633-25-5	60
		cis-13-Octadecenoic acid	142083	013126-39-1	53
44	17.627 1.72	11,13-Dimethyl-12-tetradecen-1-ol acetate	142133	1000130-81-0	76
		Docosyl propyl ether		1000406-28-2	
		Hexacosyl propyl ether	219473	1000406-28-4	50
			248661		46
45	17.775 3.42	Methyl tetratriacontyl ether	267425	1000406-30-1	90
		Octacosyl propyl ether	257323	1000406-28-5	70
		Hexacosyl propyl ether	248661	1000406-28-4	70
46	18.338 2.17	4,8,12,16-Tetramethylheptadecan-4-olide	182574	096168-15-9	72
		delta-Nonalactone			
		trans-3-Methyl-4-octanolide	29928	003301-94-8	46
			29959	039638-67-0	46
47	18.412 5.59	9-Octadecenamide, (Z)-	141029	000301-02-0	98
		9-Octadecenamide, (Z)-	141028	000301-02-0	96
		9-Octadecenamide, (Z)-	141032	000301-02-0	95
48	18.989 2.25	Stigmaterol	244153	000083-48-7	25
		6,9-Octadecadienoic acid, methyl ester	150118	056847-03-1	14
		Stigmasta-5,22-dien-3-ol, acetate(3.beta)-			
			57963	004651-48-3	12
49	19.071 2.31	Stigmaterol	244153	000083-48-7	40
		6,9-Octadecadienoic acid, methyl ester	150118	056847-03-1	14
		Stigmasta-5,22-dien-3-ol, acetate(3.beta)-			
			257963	004651-48-3	12

DISCUSSION

Bio-pesticides are effective and safer means of controlling pests, they have a mild effect on the environment compared to their synthetic counterpart, and they are specific in their target, hence preventing bioaccumulation (Kumar *et al.*, 2021; Saberi *et al.*, 2020). Bio-pesticides are made from natural substances, such as plants, microbes, and nanoparticles of biological origin, thus, making them a sustainable means of pest control (Saberi *et al.*, 2020). The effectiveness of bio-pesticides in pest management comes from various modes of action, which include actions that regulate gut disruption, pest growth, and pest metabolism. Bio-pesticides work by denaturing protein, causing metabolic disorder and paralysis, activating target-poisoning mechanisms, exhibiting multisite inhibitory actions, and releasing neuromuscular toxins and bioactive compounds (Sparks and Nauen, 2015; Dar *et al.*, 2021; Fenibo *et al.*, 2021). These multiple actions offer bio-pesticides the capacity to alter the course of pest resistance as compared to chemical pesticides. Studies have indicated that bio-pesticides are eco-friendly, possess low toxicity properties, are biodegradable, and specific in action with little or no negative impact on non-target organisms (Deravel *et al.*, 2014; Kalpana and Anil, 2021). Unlike bio-pesticides, conventional pesticides are a major source of environmental pollution, which promotes pest resistance with high post-harvest contamination and bioaccumulation in food crops (Fenibo *et al.*,

2021).

In this study, GC-FID analysis of ethanolic extract of *Chromolaena odorata* reveals the presence of phytol. Phytol has been reported to have antimicrobial activity against *Mycobacterium tuberculosis* (Rajab *et al.*, 1998; Saikia *et al.*, 2010) and *Staphylococcus aureus* (Inoue *et al.*, 2010). Therefore, siam weed might have bio-pesticidal effect. In medicinal fields, phytol has shown anti-nociceptive and antioxidant activities as well as anti-inflammatory and anti-allergic effects (Ryuet *et al.*, 2011). Recent studies have revealed that phytol is an excellent immune stimulant, superior to a number of commercial adjuvants in terms of long-term memory induction and activation of both innate and acquired immunity (Lim *et al.*, 2006). Phytol has been investigated for its potential anxiolytic, metabolism modulating, cytotoxic, antioxidant, autophagy and apoptosis-inducing, anti-nociceptive, anti-inflammatory, immune-modulating, and antimicrobial effects (Islam *et al.*, 2018). It has also been reported that phytol modulate transcription in cells via transcription factors PPAR-alpha and retinoid X receptor (RXR) (Saikia *et al.*, 2010). According to the report of Moraes *et al.*, phytol is gaining more usefulness as a food additive and in medicinal fields, as its manifest promising antischistosomal properties in vitro and in a mouse model of schistosomiasis mansoni (Moraes *et al.*, 2014).

Linoleic acid ethyl ester is an esterified form of linoleic acid. It is active against the bacteria *S. mutans*, *A. actinomycetemcomitans*, *P. gingivalis*, *S. gordonii*, and *S. sanguis* and the fungus *C. albicans* when used at a concentration of 25 µg/ml. Linoleic acid ethyl ester (3 mg/kg) prevents arteriosclerosis in rabbits fed with a high-cholesterol diet (Huang *et al.*, 2010). Linoleic acid ethyl ester has also been reported to have hypocholesterolemic, nematocidal, anti-arthritis, hepato-protective, anti-androgenic, hypocholesterolemic, 5-Alpha reductase inhibitor, anti-histaminic, anti-coronary, insecticidal, anti-eczemic, anti-acne activity (Sudha *et al.*, 2013). The mechanism of conjugated linoleic acid as insecticide is due to anti-feedant effects, increased larval mortality, decreased larval development and egg viability (Clement *et al.*, 2019). In *Saccharomyces cerevisiae*, linoleic acid performs cell toxicity by inducing apoptosis and repressing metabolic enzyme activities (Ferreira *et al.*, 2011). The very long-chain fatty acids elongases play important roles in yeast resistance to cytotoxicity induced by oleic acid through balancing the fatty acid composition in bio-membranes (Wang *et al.*, 2018). Another active ingredient of interest revealed is

stigmasterol, an important plant sterol otherwise regarded as unsaturated phytosterol occurring in plant with potential to inhibit the development of various cancerous cells by inhibiting the promotion and growth of cancer cell lines including but not limited to stomach, lung, ovary, and estrogen-dependent human breast cancers (Bradford and Awad, 2007). It is of great interest finding this extract as one of the sources of stigmasterols that is regarded as the anti-cancer therapy of the future due to its ability to scavenge cancer-causing substances (Lim *et al.*, 2006).

Copaene (COP) is a tricyclic sesquiterpenes derived from different plants; leaves (Afoulouset *al.*, 2013) strongly attractive to an agricultural pest. As a sesquiterpenes it has biological activities such as antimicrobial (Wu *et al.*, 2012), antibacterial (Stojanović-Radić *et al.*, 2012), antioxidant, anti-fungi (Al-maskriet *al.*, 2013). Hexadecanoic acid, ethyl ester is a palmitic acid ethyl ester, it has been reported to have antioxidant, hypocholesterolemic, nematocide, pesticide, anti-androgenic flavor, hemolytic, aldehyde reductase inhibitor activity (Sudha *et al.*, 2013). 1H-Pyrazole, 4,5-dihydro-3-methyl-1-propyl- with ratio (14.833%) as antibacterial properties (Gottenbos *et al.*, 2002). Caryophyllene oxide is an oxygenated sesquiterpenoid, well-recognized as a protective compound in nutrition, medications and cosmetics. Caryophyllene oxide has some pharmacological potentials like anticholinesterase, analgesic, anti-inflammatory, antifungal activities (Park *et al.*, 2011). Caryophyllene oxide (CPO) is a sesquiterpene isolated from essential oils of medicinal plants such as clove (*Eugenia caryophyllata*), black pepper (*Piper nigrum* L.), oregano (*Origanum vulgare* L.), guava (*Psidium guajava*) and cinnamon (*Cinnamomum* spp.), and act as a strong anticancer agent also can stop the proliferation of a wide variety of tumor cells, consisting of breast, lung, liver, and prostate adenocarcinoma as reported in several studies (Park *et al.*, 2011).

Vanillic acid (VA) (4-hydroxy-3-methoxybenzoic acid) is a dietetic phenolic composite present in plants and fruits. VA is a metabolic formation of caffeic acid that leads to the development of urine in an inhabitant. Consequent to intake of coffee, chocolate or green tea, and it can demonstrate antioxidant, antimicrobial, anti-genotoxic, and antitumor activities. VA can suppress hepatic fibrosis in chronic liver injury, and it also exhibits chemo-preventive effects in experimentally induced carcinogenesis in rats. 1,2,3-Trimethoxybenzene (TMB) is an organic compound containing three methoxy groups in benzene ring having a molecular formula $C_9H_{12}O_3$

and molecular weight of 168.19. These compounds can be used in pharmaceuticals (analgesics, antiseptics), biocides, formulating insect attractants, flavoring agents. They are also useful in the preparation of antioxidants, stabilizers, UV absorbers for plastics and rubbers.

CONCLUSION

In conclusion, analysis of *Chromolaena odorata* has shown its bio-pesticide composition. The chemical composition of bio-pesticides is diverse, comprising various natural ingredients. They offer environmentally friendly pest control options with reduced risks to non-target organisms. While bio-pesticides have specific target pest control and are biodegradable, they may require frequent application and face limitations in persistence and efficacy. Overall, bio-pesticides play a valuable role in integrated pest management strategies, promoting sustainable agricultural practices and reducing reliance on synthetic pesticides.

REFERENCES

- Aba, P.E., Joshua, P.E., Ezeonugu, F.C., Ezeja, M.I., Omoja, V.U., Umeakuana, P.U. (2015). Possible anti-diarrhoeal potential of ethanol leaf extract of *Chromolaena odorata* in castor oil-induced rats. *J. Complement. Integr. Med.* 12:301–306.
- Adedapo, A.A., Oyagbemi, A.A., Fagbohun, O.A., Omobowale, T.O., Yakubu, M.A. (2016). Evaluation of the anticancer properties of the methanol leaf extract of *Chromolaena odorata* on HT29 lung cancer cell line. *The FASEB Journal* 30(Supplement):1193–1196.
- Afoulous, S., Ferhout, H., Raelison, E.G., Valentin, A., Moukarzel, B., Couderc, F., Bouajila, J. (2013). Chemical composition and anticancer, anti-inflammatory, antioxidant and antimalarial activities of leaves essential oil of *Cedrelopsis grevei*. *Food Chem. Toxicol.* 56:352–362.
- Airaodion, A.I., Ogbuagu, E.O., Ogbuagu, U., Awosanya, O.O., Airaodion, E.O. (2019). Hypoglycemic and hypolipidaemic effect of methanolic extract of *Corchorus olitorius* leaves in Wistar rats. *Asian Plant Research Journal*. 7(6):1–12.
- Alisi, C.S., Onyeze, G.O., Ojako, O.A., Osuagwu, C.G. (2011). Evaluation of the protective potential of *Chromolaena odorata* Linn. extract on carbon tetrachloride-induced oxidative liver damage. *Int. J. Biochem. Res. Rev.* 1:69–81.
- Al-maskri, A.Y., Hanif, M.A., Al-Maskari, M.Y., Abraham, A.S., Al-sabahi, J.N., Al-Mantheri, O. (2013). Essential oil from *Ocimum*

- basilicum* (Omani Basil): a desert crop. *Nat. Prod. Commun.* 6:1487–1490.
- Amazu, L.U., Omeregbe, P., Ajugwo, A.O., Ifezulike, C.C., Azikiwe, C.C. (2013). Anticonvulsant potency of the leaf extract of *Chromolaena odorata* in rats. *Unique Res. J. Med. Sci.* 1:64–69.
- AOAC. (2019). *Official Methods of Food Analysis*. Association of Official Analytical Chemists, Washington DC, USA. 16th Edition.
- Asomugha, R.N., Okafor, P.N., Ijeh, I.I., Orisakwe, O.E., Asomugha, A.L. (2014). Hepatic effects of aqueous extract of *Chromolaena odorata* in male Wistar albino rats. *Pharmacol. Online*. 1:127–136.
- Atindehou, M., Lagnika, L., Guerold, B., Strub, J.M., Zhao, M., Dorsselaer, A.V. (2013). Isolation and identification of two antibacterial agents from *Chromolaena odorata* L. active against four diarrheal strains. *Adv. Microbiol.* 3:115–121.
- Boudjeko, T., Megnekou, R., Woguia, A.L., Kegne, F.M., Ngomoyogoli, J.E., Tchoum, C. D. (2015). Antioxidant and immunomodulatory properties of polysaccharides from *Allanblackia floribunda* stem bark and *Chromolaena odorata* leaves. *BMC Res. Notes*. 8:759.
- Bouda, H., Tapondjou, L.A., Fontem, D.A., Gumedzoe, M.Y.D. (2001). Effect of essential oils from leaves of *Ageratum conyzoides*, *Lantana camara* and *Chromolaena odorata* on the mortality of *Sitophilus zeamais*. *J. Stored Prod. Res.* 2(1):103–109.
- Bradford, P.G., Awad, A.B. (2007). Phytosterols as anticancer compounds. *Mol. Nutr. Food Res.* 51:161–170.
- Chavan, M.J., Wakte, P.S., Shinde, D.B. (2010). Analgesic and anti-inflammatory activity of caryophyllene oxide from *Annona squamosa* L. bark. *Phytomedicine*. 17:149–151.
- Clements, J., Groves, R.L., Cava, J., Barry, C.C., Chapman, S., Olson, J.M. (2019). Conjugated linoleic acid as a novel insecticide targeting the agricultural pest *Leptinotarsa decemlineata*. *PLoS ONE*. 14(11):e0220830.
- Dar, S.A., Wani, S.H., Mir, M., Showkat, A., Dolkar, T., Dawa, T. (2021). Biopesticides: mode of action, efficacy and scope in pest management. *J. Adv. Res. Biochem. Pharmacol.* 4:1–8.
- Deravel, J., Krier, F., Jacques, P. (2014). Biopesticides, a complementary and alternative approach to the use of agrochemicals: A review. *Biotechnologie, Agronomie, Société et Environnement*. 18:220–232.
- De-Moraes, J., de-Oliveira, R.N., Costa, J.P., Junior, A.L.G., de-Sousa, D.P., Freitas, R.M. (2014). Phytol, a diterpene alcohol from chlorophyll, as a drug against neglected tropical disease *Schistosomiasis mansoni*. *PLoS Negl. Trop. Dis.* 8(1):2617.
- Ezenyi, I.C., Salawu, O.A., Kulkarni, R., Emeje, M. (2014). Antiplasmodial activity-aided isolation and identification of quercetin-4'-methyl ether in *Chromolaena odorata* leaf fraction with high activity against chloroquine-resistant *Plasmodium falciparum*. *Parasitol. Res.* 113:4415–4422.
- Félicien, A., Alitonou, G.A., Djenontin, T.S., Tchobo, F., Yèhouénou, B., Chantal, M., Sohounhloué, D. (2012). Chemical composition and biological activities of the essential oil extracted from the fresh leaves of *Chromolaena odorata* growing in Benin. *ISCAJ. Biol. Sci.* 1(3):7–13.
- Fenibo, E.O., Ijoma, G.N., Matambo, T. (2021). Biopesticides in sustainable agriculture: A critical sustainable development driver governed by green chemistry principles. *Front. Sustain. Food Syst.* 5:619658.
- Ferreira, T.C., de Moraes, L.M.P., Campos, E.G. (2011). Cell density-dependent linoleic acid toxicity to *Saccharomyces cerevisiae*. *FEMS Yeast Res.* 11:408–417.
- Gautier, L. (1992). Taxonomy and distribution of a tropical weed, *Chromolaena odorata* (L.) R. King and H. Robinson. *Candollea*. 47:645–662.
- Gottenbos, B., Busscher, H.J., van der Mei, H.C., Nieuwenhuis, P. (2002). Pathogenesis and prevention of biomaterial centered infections. *J. Mater. Sci.: Mater. Med.* 13(8):717–722.
- Huang, C.B., George, B., Ebersole, J.L. (2010). Antimicrobial activity of n-6, n-7 and n-9 fatty acids and their esters for oral microorganisms. *Arch. Oral Biol.* 55(8):555–560.
- Inoue, Y., Hada, T., Shiraishi, A., Hirose, K., Hamashima, H. (2005). Biphasic effects of geranylgeraniol, teprenone, and phytol on the growth of *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* 49:1770–1774.
- Islam, M.T., Ali, E.S., Uddin, S.J., et al. (2018). Phytol: A review of biomedical activities. *Food Chem. Toxicol.* 121:82–94.
- Joshi, R.K. (2013). Chemical composition of the essential oils of aerial parts and flowers of *Chromolaena odorata*. *J. Essent. Oil Bear. Plants*. 16(1):71–75.
- Kalpana, T., Anil, T. (2021). A review of biopesticides and their plant phytochemicals information. *Ann. Romanian Soc. Cell Biol.* 25:3576–3588.
- Koba, K., Nénonéné, A.Y., Catherine, G., et al. (2011). Chemical composition and cytotoxic activity of essential oil of *Chromolaena odorata* L. growing in Togo. *J. Essent. Oil*

- Bear. Plants.* 14(4):423–439.
- Kossouh, C., Moudachirou, M., Adjakidje, V., *et al.* (2011). Volatile constituents of *Chromolaena odorata* leaves from Benin. *J. Essent. Oil Bear. Plants.* 14(2):224–228.
- Kouame, P.B., Jacques, C., Bedi, G., *et al.* (2013). Phytochemicals isolated from leaves of *Chromolaena odorata*: Impact on viability and clonogenicity of cancer cell lines. *Phytother. Res.* 27:835–840.
- Kumar, J., Ramlal, A., Mallick, D., Mishra, V. (2021). An overview of some biopesticides and their importance in plant protection for commercial acceptance. *Plants.* 10:1185.
- Lim, S.Y., Meyer, M., Kjonaas, R.A., Ghosh, S.K. (2006). Phytol-based novel adjuvants in vaccine formulation. *J. Immune Based Ther. Vaccines.* 4:6.
- Naidoo, K.K., Coopoomsamy, R.M., Naidoo, G. (2011). Screening of *Chromolaena odorata* for antibacterial and antifungal properties. *J. Med. Plant Res.* 5:4859–4862.
- Ngono, N.A., Ebelle Etame, R., Ndifor, F., *et al.* (2006). Antifungal activity of *Chromolaena odorata*. *Chemotherapy.* 52:103–106.
- Noudogbessi, J.P., Kossou, D., Sohounhloué, D.C.K. (2008). Effet insecticide, ovicide et larvicide des huiles essentielles de *Chromolaena odorata*. *J. Soc. Ouest-Afr. Chim.* 26(1):41–51.
- Okafor, C.S., Ononuju, C.O., Eze, C.G., Nwosu, O.K., Ifedilichukwu, N.H. (2019). *Chromolaena odorata* (Siam weed) as biopesticide against beans weevils. *Agric. Biol. Sci. J.* 5(3):105–109.
- Omokhua, A.G., McGaw, L.J., Finnie, J.F., Van Staden, J. (2016). *Chromolaena odorata* in sub-Saharan Africa: A synthesis and review of its medicinal potential. *J. Ethnopharmacol.* 183:112–122.
- Onkaramurthy, M., Veerapur, V.P., Thippeswamy, B.S., *et al.* (2013). Anti-diabetic and anti-cataract effects of *Chromolaena odorata*. *J. Ethnopharmacol.* 145:363–372.
- Owolabi, M.S., Ogundajo, A., Yusuf, K.O., *et al.* (2010). Chemical composition and bioactivity of the essential oil of *Chromolaena odorata* from Nigeria. *Rec. Nat. Prod.* 4(1):72–78.
- Owoyele, V.B., Adediji, J.O., Soladoye, A.O. (2005). Anti-inflammatory activity of aqueous leaf extract of *Chromolaena odorata*. *Inflammopharmacology.* 13:479–484.
- Pamo, E.T., Amvam Zollo, P.H., Tendonkeng, F., *et al.* (2004). Composition chimique et effet acaricide des huiles essentielles des feuilles de *Chromolaena odorata*. *Livestock Res. Rural Dev.*
- Pandith, H., Zhang, X., Liggett, J., Min, K.W., Gritsanapan, W., Baek, S.J. (2013). Hemostatic and wound healing properties of *Chromolaena odorata* leaf extract. *ISRN Dermatol.*
- Pandith, H., Zhang, X., Thongpraditchote, S., Wongkrajang, Y., Gritsanapan, W., Baek, S.J. (2013). Effect of Siam weed extract on anti-inflammatory activity through NF- κ B pathway. *J. Ethnopharmacol.* 147:434–441.
- Pandurangan, A., Rana, K., Singh, A. (2015). Evaluation wound healing activity of leaves of *Chromolaena odorata* Linn. *Int. J. Pharm. Sci. Lett.* 5:555–557.
- Park, K.R., Nam, D., Yun, H.M. (2011). β -Caryophyllene oxide inhibits growth and induces apoptosis. *Cancer Lett.* 312:178–188.
- Phan, T.T., Wang, L., See, P., *et al.* (2001). Phenolic compounds of *Chromolaena odorata* protect cultured skin cells from oxidative damage. *Biol. Pharm. Bull.* 24:1373–1379.
- Rajab, M.S., Cantrell, C.L., Franzblau, S.G., Fischer, N.H. (1998). Antimycobacterial activity of (E)-phytol and derivatives. *Planta Med.* 64:2–4.
- Rouw, A. (1991). The invasion of *Chromolaena odorata* and competition with native flora in Côte d'Ivoire. *J. Biogeogr.* 18:13–23.
- Ryu, K.R., Choi, J.Y., Chung, S., Kim, D.H. (2011). Anti-scratching behavioral effect of phytol. *Planta Med.* 77:22–26.
- Saberi, F., Marzban, R., Ardjmand, M., Shariati, F.P., Tavakoli, O. (2020). Optimization of culture media to enhance the ability of local *Bacillus thuringiensis* var. *tenebrionis*. *J. Saudi Soc. Agric. Sci.* 19:468–475.
- Saikia, D., Parihar, S., Chanda, D., Ojha, S., Kumar, J.K. (2010). Antitubercular potential of some semisynthetic analogues of phytol. *Bioorg. Med. Chem. Lett.* 20:508–512.
- Santos, C.C.M.P., Salvadori, M.S., Mota, V.G., *et al.* (2013). Antinociceptive and antioxidant activities of phytol. *Neurosci. J.* Article ID 949452.
- Sparks, T.C., Nauen, R. (2015). IRAC: Mode of action classification and insecticide resistance management. *Pest Biochem. Physiol.* 121:122–128.
- Stanley, M.C., Ifeanyi, O.E., Nwakaego, C.C., Esther, I.O. (2014). Antimicrobial effects of *Chromolaena odorata* on some human pathogens. *Int. J. Curr. Microbiol. Appl. Sci.* 3:1006–1012.
- Stojanović-Radić, Z., Comić, L., Radulović, N., *et al.* (2012). Antistaphylococcal activity of *Inula helenium* root essential oil. *Eur. J. Clin. Microbiol. Infect. Dis.* 31:1015–1025.
- Sudha, T., Chidambarampillai, S., Mohan, V. (2013). GC-MS analysis of bioactive components of *Fluggea leucopyrus*. *J. Appl. Pharm. Sci.* 3(5):126–130.

- Suksamrarn, A., Chotipong, A., Suavansri, T., *et al.* (2004). Antimycobacterial activity and cytotoxicity of flavonoids from *Chromolaena odorata*. *Arch. Pharm. Res.* 27:507–511.
- Suwaibah, M., Zatilfarihiyah, R., Aslizahi, M.A., Baharum, S.N. (2012). Chemical composition of local *Eupatorium odoratum* essential oil using GC-MS. *UMT 11th International Annual Symposium on Sustainability Science and Management*, 1383–1388.
- Thang, P.T., Patrick, S., Teik, L.S., Yung, C.S. (2001). Antioxidant effects of *Chromolaena odorata* leaf extracts. *Burns*. 27:319–327.
- Thoden, T.C., Boppré, M., Hallmann, J. (2007). Pyrrolizidine alkaloids of *Chromolaena odorata* act as nematocidal agents. *Nematology*. 9:343–349.
- Uhegbu, F.O., Imo, C., Onwuegbuchulam, C.H. (2016). Lipid lowering, hypoglycemic and antioxidant activities of *Chromolaena odorata*. *J. Med. Plants Stud.* 4:155–159.
- Vijayaraghavan, K., Mohamed Ali, S., Maruthi, R. (2013). Phytochemical screening and antioxidant activity of *Chromolaena odorata*. *Int. J. Innov. Res. Sci. Eng. Technol.* 2:7315–7321.
- Wang, Q., Du, X., Ma, K., *et al.* (2018). A critical role for very long-chain fatty acid elongases in oleic acid-mediated *Saccharomyces cerevisiae* cytotoxicity. *Microbiol. Res.* 207:1–7.
- Wu, Y.X., Chen, Y.J., Liu, C.M., Gao, K. (2012). Four new sesquiterpenoids from *Ligularia cymbulifera*. *J. Asian Nat. Prod. Res.* 14:1130–1136.